

## Crystallization and preliminary crystallographic studies of pink color chromoprotein from *Pleurotus salmoneostramineus* L. Vass.

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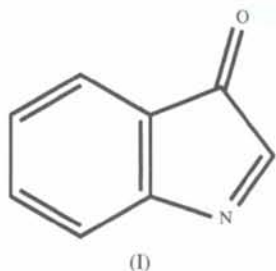
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### Abstract

A chromoprotein from *Pleurotus salmoneostramineus* L. Vass. has been purified and crystallized. The needle-shaped crystal has monoclinic space group *C2* with the cell dimensions of  $a = 118.5$ ,  $b = 59.7$ ,  $c = 31.8$  Å and  $\beta = 114^\circ$ . The crystal diffracts to 1.8 Å resolution with a synchrotron radiation X-ray source.

### 1. Introduction

*Pleurotus salmoneostramineus* L. Vass., a type of mushroom found in Siberia, Japan and New Guinea, is known for its beautiful pink color. This mushroom includes a pink chromoprotein with an absorption spectrum maximum at 496 nm. This protein has a pigment molecule, 3*H*-indol-3-one (I) which occupies a part of the active site (Takekuma, Takekuma, Matsubara, Inaba & Yoshida, 1994) and has three kinds of metals, Zn, Fe, and Cu. 3*H*-indol-3-one has an absorption maximum at 456 nm in methanol and a bathochromic shift occurs in the chromoprotein. Takekuma *et al.* have reported that the protein produces oxygen molecules from water on light irradiation. This phenomenon implies that



this mushroom has a photosynthetic function in which the chromoprotein plays an important role (Takekuma *et al.*, 1994). The aim of the present study is to reveal the structure–function relationship of the chromoprotein with the novel photosynthetic mechanism.

### 2. Purification and crystallization

Chromoprotein was extracted by soaking the mushrooms in water. Crude chromoprotein was freeze-dried and stored at 195 K. The protein (0.5 g) was dissolved in 1 ml of Tris–HCl buffer (pH 7.0), then applied onto a Sephacryl S-300HR column (Pharmacia, 3 × 80 cm, pre-equilibrated in Tris–HCl buffer, pH 7.0). The fractions containing the chromoprotein were collected and dialyzed for 12 h against glycine–NaOH

buffer (pH 10.0). It was loaded on DEAE–Sephacryl column (2 × 30 cm) equilibrated in glycine–NaOH buffer (pH 10.0) and then washed with 20 mM NaCl solution in the elution buffer. The purified protein was dialyzed against water and concentrated up to 10 A cm<sup>-1</sup> at 496 nm.

Single crystals were obtained by the vapor-diffusion method. The reservoir consists of 50 mM buffer solution (Tris–HCl, pH 9.0) containing 21% (w/v) polyethylene glycol 4600 (Sigma) and 200 mM sodium formate. The protein drop was prepared by mixing 3 μl of protein solution with 3 μl of reservoir solution. Single red-colored crystals appeared as prismatic needles within one week (Fig. 1).

### 3. X-ray analysis

Preliminary X-ray experiments were carried out on a Rigaku R-AXIS IIC imaging-plate detector system equipped with a Rigaku RU-300 rotating-anode X-ray generator (fine-focused Cu *K*α, operating at 40 kV and 100 mA). The Laue symmetry and cell dimensions were determined by the *PROCESS* program package (Higashi, 1989; Sato *et al.*, 1992) and systematic absences were confirmed by pseudo-precession pictures using the program *HKLPLOT* (Eleanor Dodson, unpublished data; Collaborative Computational Project, Number 4, 1994). The crystals belong to monoclinic space group *C2* with the unit-cell dimensions  $a = 118.5$ ,  $b = 59.7$ ,  $c = 31.8$  Å and  $\beta = 114^\circ$ . Supposing one molecule per asymmetric unit,  $V_m$  value is calculated to be 2.1 Å<sup>3</sup> Da<sup>-1</sup> (Matthews, 1968). Diffraction was found on imaging plates up to 2.5 Å resolution.

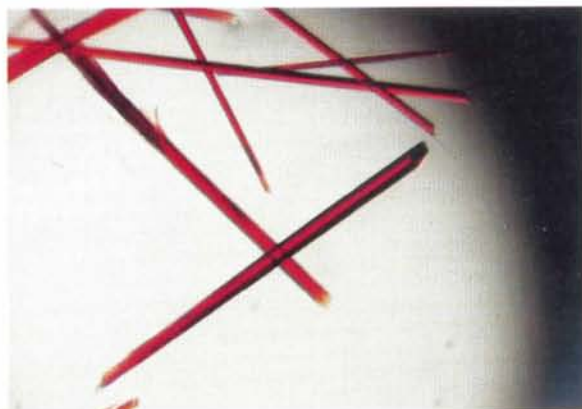


Fig. 1. Crystals of chromoprotein from *Pleurotus salmoneostramineus* L. Vass. The biggest crystal is approximately 0.5 × 0.1 × 0.07 mm.

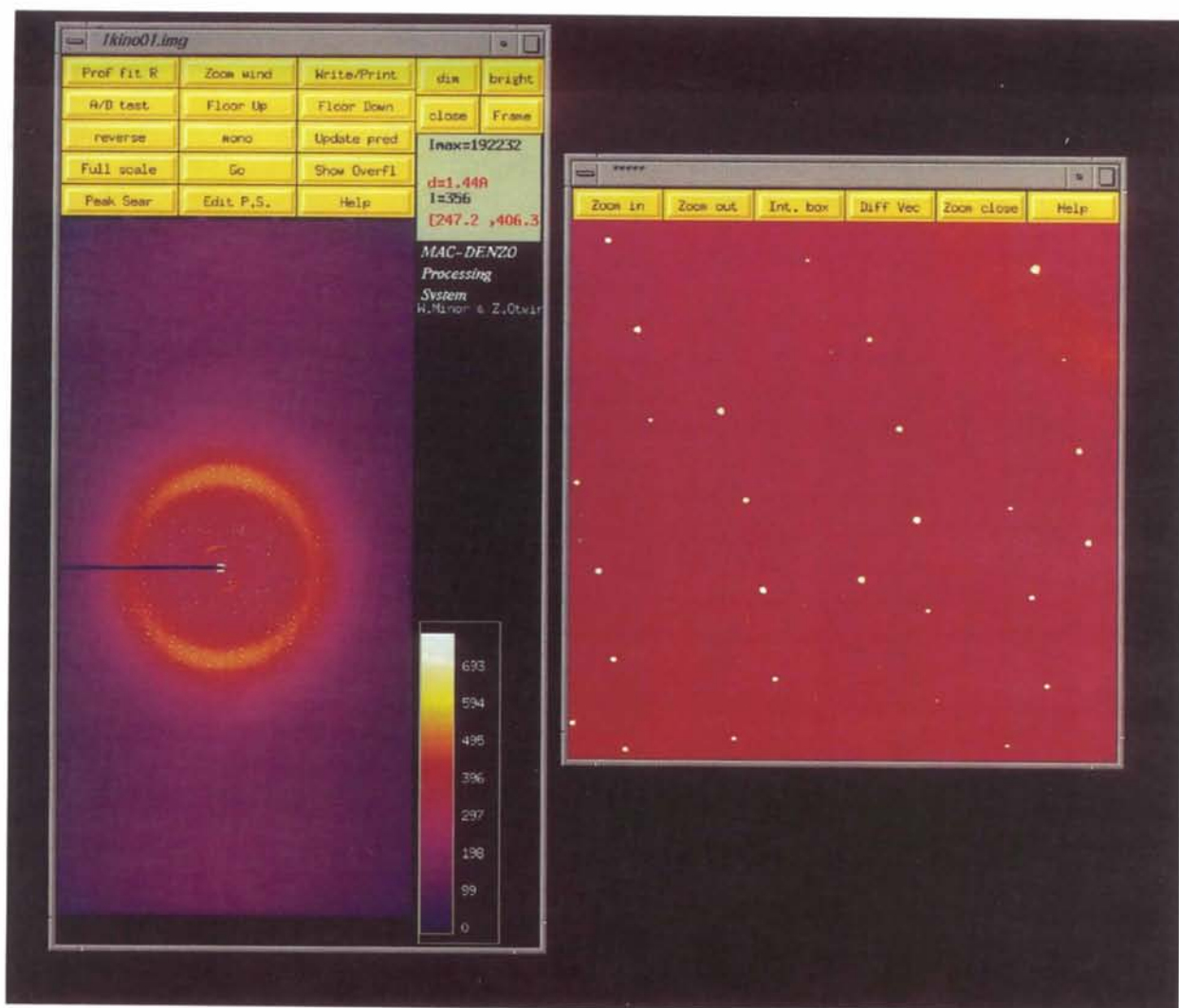


Fig. 2. A Weissenberg diffraction pattern from a chromoprotein crystal (rotation axis  $b$ ). This image was obtained with a Weissenberg camera for macromolecules (Sakabe, 1983; cassette distance and radius are 430 mm) equipped with an imaging plate ( $40 \times 80$  cm).

Diffraction intensity data were collected by using synchrotron radiation (BL-18B beamline, Photon Factory, KEK, Japan). The Weissenberg camera for macromolecules (Sakabe, 1983) and large imaging plates ( $40 \times 80$  cm) were used for data collection. The crystal diffracts up to  $1.8 \text{ \AA}$  resolution and 15 frames with a rotation angle of  $12.5^\circ$  for each frame were stored (Fig. 2). During the data collection no obvious radiation damage was detected. Data processing was carried out by the programs *DENZO* and *SCALEPACK* (Otwinowski, 1993). The combined set gave 71 241 reflections to  $1.8 \text{ \AA}$  resolution in total, which were reduced to 18 198 unique reflections with an  $R_{\text{merge}}$  of 4.8% ( $R_{\text{merge}} = \sum |I - \langle I \rangle| / \sum I$ ) and the completeness of 85.7% (61.4% for  $1.80\text{--}1.86 \text{ \AA}$ ).

A search for heavy-atom derivatives is currently under way for structure determination by isomorphous replacement method.

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